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Inhibition of atherosclerosis by the serine palmitoyl transferase inhibitor myriocin is associated with reduced plasma glycosphingolipid concentration

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ABSTRACT

Glycosphingolipids (GSL) have been implicated as potential atherogenic lipids. Inhibition of hepatic serine palmitoyl transferase (SPT) reduces plasma sphingomyelin (SM) levels in the absence of changes in cholesterol or triglyceride (TG) concentration and this leads to a reduction of atherosclerosis in a polipoprotein-E gene knockout (apo $E^{-/-}$) mice. The possibility that the reduced atherosclerosis resulting from SPT inhibition is associated with decreases in plasma GSL concentration has not been examined and was the primary aim of this investigation. We show that intraperitoneal delivery of the SPT inhibitor myriocin for 9 weeks inhibits atherosclerosis in apo $E^{-/-}$ mice fed a high fat diet. Lesion inhibition was most pronounced at the aortic arch and distal sites of the thoracic and abdominal aorta. There was also a trend towards a reduction in lesion area at the aortic root. Myriocin treatment resulted in significant reductions in both plasma SM and GSL concentration of 42% and 25%, as assessed by enzymatic and HPLC methods, respectively. Moreover, SM and GSL concentrations were significantly correlated, indicating that SPT inhibition suppresses the synthesis of both these sphingolipids concomitantly. The inhibition of atherosclerosis induced by myriocin was not associated with changes in plasma cholesterol or TG concentrations or lipoprotein profiles as determined by FPLC. These data indicate that therapeutic reduction of plasma SM and/or GSL concentrations may offer a novel treatment for atherosclerosis.

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1. Introduction

Atherosclerosis is a major cause of cardiovascular disease (CVD) and accounts for \sim 50% of all deaths in westernised countries [1]. Atherosclerosis develops as a consequence of

multiple pathways that involve inflammation, oxidative stress, dysregulated cellular proliferation and lipid accumulation [2–4]. The accumulation of lipids, including cholesterol, sphingomyelin (SM) and glycosphingolipids (GSL), in atherosclerotic lesions is well known [5–7]. Plasma cholesterol levels

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are widely used as a predictor of CVD risk and plasma SM and GSL concentrations are also associated with atherosclerosis risk [8-10]. In the case of GSL, potential pro-atherogenic properties have been proposed. For example, lactosylceramide (LacCer) promotes cholesterol accumulation in macrophage foam-cells [11], inhibits cellular cholesterol removal via the ABCA1/apoA-I pathway [12], induces monocyte adhesion to endothelial cells [13] and stimulates vascular smooth muscle cell proliferation [14]. Other studies have reported that ganglioside GM3 accelerates low-density lipoprotein (LDL) uptake by macrophages which results in the generation of lipid-laden foam cells [7]. Furthermore, GM3 and GD3 promote the adhesion of platelets to sites of atherosclerotic lesion formation and GD3 stimulates production of reactive oxygen species, regulates smooth muscle cell phenotype and inhibits metalloproteinase-9 expression; all events potentially contributing to plaque instability and atherosclerosis [15,16]. In more general terms, vascular accumulation of GSL could impact on atherogenesis via regulation of cellular signalling, activation, recognition, differentiation, fibrinolytic activity, nitric oxide production, and response to growth factors (see [10,17]).

We have begun to assess whether suppression of GSL synthesis in vitro has any impact on cellular pathways that could be considered as anti-atherogenic. One potential pathway involves the removal of cholesterol from macrophages via ATP-binding cassette transporter A1 (ABCA1) in a process known as reverse cholesterol transport [18,19]. Macrophages in atherosclerotic lesions are derived from circulating monocytes that infiltrate the endothelium and become lodged in the intima where they endocytose modified low density lipoproteins (mLDLs, e.g. modified by proteoglycans, enzymes or oxidants) and become cholesterol-loaded foam cells [1]. We have shown that inhibition of macrophage foam cell GSL synthesis using the glucosylceramide synthase inhibitor Dthreo-1-phenyl-2-decanoylamino-3-morpholino-1-propanol (PDMP) stimulated cholesterol efflux [12]. Although this work suggests that GSL synthesis inhibition may afford protection in the context of atherosclerosis development, the key question as to whether modulation of GSL levels in vivo has an impact on atherosclerosis remains unanswered.

As is the case in humans, plasma and aortic GSL levels were found to be increased with atherosclerotic lesion development in apoE-/- mice, thereby identifying an appropriate in vivo model to study the role of GSL in atherogenesis [10]. Subsequent research showed that increasing vascular GSL burden in apoE^{-/-} mice by crossing with α -galactosidase deficient mice (the latter mimic the GSL storage disorder known as Fabry disease) resulted in accelerated atherosclerosis [20]. This finding therefore provided further support to the view that GSL may be pro-atherogenic. Other recent work from two independent groups has shown that inhibition of serine palmitoyl transferase (SPT), the enzyme catalysing the first step in the sphingolipid biosynthetic pathway, dramatically reduced atherosclerotic lesion formation in apoE^{-/-} mice [21,22]. The aim of these studies was in fact to assess the potential impact of SM depletion on atherogenesis and the rationale was based on (as noted above) the observation that plasma SM levels are positively correlated with atherosclerosis risk [9]. In light of the original studies by Miyake et al. reporting that inhibition of SPT with myriocin suppresses GSL synthesis in vitro [23], we speculated that the inhibition of atherosclerosis observed in myriocin-treated apo $E^{-/-}$ mice may not be exclusively due to inhibition of SM synthesis but also to reduced GSL synthesis [24]. The aim of the present study was therefore to examine whether myriocin-mediated inhibition of atherosclerosis in apo $E^{-/-}$ mice was associated with a reduced concentration of plasma GSL.

2. Materials and methods

2.1. Materials

All organic solvents were analytical grade and purchased from Merck (Darmsdadt, Germany). Purified leech (Macrobdella decora) ceramide glycanase (E.C.3.2.1.123) was from V-Labs (Covington, LA) and myriocin, (2S,3R,4R,6E)-2-amino-3,4-dihydroxy-2-(hydroxymethyl)-14-oxo-6-eicosenoic acid, from Sigma (Castle Hill, NSW, Australia). All other reagents were of the highest purity available and purchased through standard commercial suppliers.

2.2. Animals and diet

Male apoE $^{-/-}$ mice were supplied by the Animal Resources Centre (Canning Vale, WA, Australia). Mice were fed standard chow until 10 weeks of age then changed to a high fat diet containing 22% (w/w) fat and 0.15% (w/w) cholesterol (Diet No. SF00-219, Specialty Feeds, Glen Forest, WA, Australia). At the same time the mice were changed to a high fat diet, myriocin or vehicle control administration commenced as described previously [22]. In brief, mice were injected intra-peritoneally (i.p.) with filter sterilized (0.2 μ m) myriocin dissolved in phosphate-buffered saline (PBS) or with PBS for controls (n = 7 in each group). Injection volume for all mice was 100 μ l and myriocin dose was of 0.3 mg/kg every 48 h.

2.3. Assessment of atherosclerotic lesions

After 9 weeks on the high fat diet \pm myriocin treatment, mice were fasted overnight, euthanased, plasma collected and perfusion fixed hearts and aortae dissected and the root, arch, and descending thoracic (at the branch point of third intercostal pair) and abdominal (at the coeliac branch point) aortic sections prepared for assessment of lesion area based on previous methods [25,26] as described in detail [27]. Morphometric data was collected for all four sites after sections were subjected to Verhoeff staining.

2.4. Plasma lipid analysis

Plasma cholesterol, TG and SM analysis was by enzymatic methods [10,28]. Plasma GSL analysis was by normal phase HPLC [10,29] with minor modifications. In the present study, 0.1 U ceramide glycanase was used to hydrolyse total neutral and charged GSL in 40 μl plasma and the resultant GSL-derived glycans fluorescently labelled with 2-aminobenzamide (2-AB) and analysed as a single sample (rather than analysing neutral and charged glycans separately).

2.5. Lipoprotein sub-fractionation

Fresh plasma (200 μ l, stored at 4 °C < 48 h) from 2 mice (1 control and 1 myriocin-treated) was mixed with 200 μ l 20 mM sodium phosphate buffer, pH 7.8, and this diluted plasma subjected to FPLC (Pharmacia AKTAexplorer 100, Uppsala, Sweden) using a Superose-6 and Superose-12 column (10 mm \times 370 mm each) in series as described previously [30]. The columns were eluted at a flow rate of 0.25 ml/min with 20 mM sodium phosphate buffer, pH 7.8, and the eluant was monitored at 280 nm.

2.6. Statistical analysis

All data are presented as means \pm S.E. Statistical significance for differences in lesion areas and plasma lipid concentrations was determined using the Mann–Whitney U test and Student's

t-test, respectively. Difference were considered significant where P < 0.05.

3. Results

In order to confirm that myriocin inhibits atherosclerosis in apoE^{-/-} mice fed a high fat diet we used i.p. administration at a dose of 0.3 mg/kg every 48 h for 9 weeks and assessed lesion area in the proximal and distal aorta. In control animals, injected with PBS, extensive lesions developed at all four sites examined. Lesions were most advanced at the aortic root with complex lesions containing necrotic regions and cholesterol crystals clearly detected (Fig. 1). Similarly, significant lesions were present at the inner curvature of the aortic arch and at the proximal branch points of the arch as well as in the thoracic and abdominal aortic segments at the third intercostal and coeliac

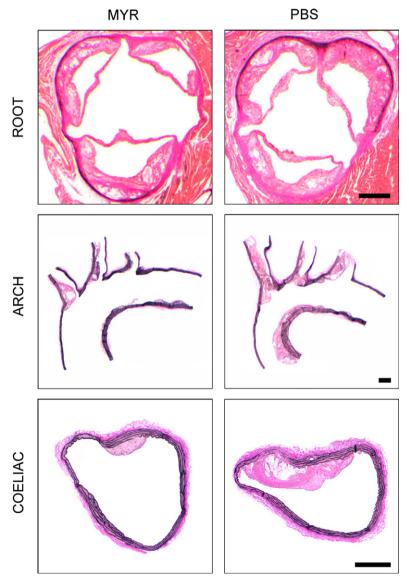


Fig. 1 – Impact of myriocin on atherosclerosis development at the aortic root, arch and coeliac branch point. Apo $E^{-/-}$ mice were fed a high fat diet for 63 days and concurrently treated i.p. with myriocin (MYR) at 0.3 mg/kg/48 h or phosphate buffered saline (PBS) as vehicle control. Lesions were assessed at the aortic root (ROOT), arch (ARCH) and coeliac branch point (COELIAC). Representative sections are shown after Verhoeff staining. Scale bar = 250 μ m.

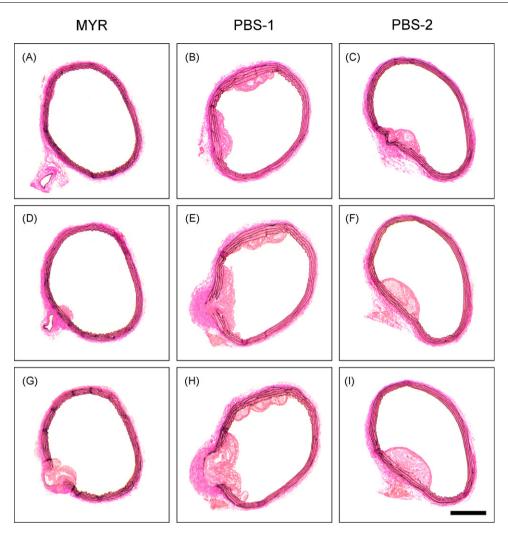


Fig. 2 – Impact of myriocin on atherosclerosis development at the third intercostal branch point of the thoracic aorta. Apo $E^{-/-}$ mice were fed a high fat diet for 63 days and concurrently treated i.p. with myriocin (MYR) at 0.3 mg/kg/48 h i.p. or phosphate buffered saline (PBS-1 and PBS-2) as vehicle control. Six lesions sections were cut at 100 μ m steps both above and below the branch point. For "MYR" and "PBS-1", sections are shown at 200 μ M (A and B) and 100 μ M (D and E) above the branch and at the branch (G and H). For "PBS-2", sections are shown just below the branch (C) and at 100 μ M (F) and 200 μ M (I) below the branch point. Representative sections are shown after Verhoeff staining. Scale bar = 250 μ m.

branches, respectively (Figs. 1 and 2). There was a trend towards reduced lesion size at the root in the myriocin treated mice, however, this did not reach statistical significance (Fig. 3). At all three of the remaining sites, myriocin significantly reduced lesion development and this was particularly noticeable at the third intercostal and coeliac branch points where average lesion area was reduced by 69% and 64%, respectively (Fig. 3).

Lesion morphology at the aortic root, arch and coeliac branch points was uniform in most of the control animals (data not shown), whereas variability was noted at the third intercostal branch point (e.g. Fig. 3). This variation is demonstrated by lesion morphology that ranged from highly focused lipid-rich lesions with a well defined fibrous cap that extended well into the arterial lumen (Fig. 2, PBS-2) to a broader, thinner lesion that extend around the inner lumenal surface (Fig. 2, PBS-1). It was clear that despite the differences in lesion morphology observed at this site, cross-sectional

areas were similar in the control animals and myriocin treatment significantly inhibited lesion development (Fig. 3).

Previous studies have shown that oral administration of myriocin (0.3 mg/kg/day) reduced apoE^{-/-} mouse plasma cholesterol and TG concentrations by 74% and 47%, respectively, and this was associated with major reductions in both very low density lipoprotein (VLDL) and low density lipoprotein (LDL) lipoprotein fractions and a doubling of high density lipoprotein (HDL) level [31]. In contrast, i.p. administration of myriocin (using the same approach as we have here) reduced liver SPT activity by 50% and reportedly had no significant impact on either plasma cholesterol or TG concentration or on lipoprotein profile [22]. We also assessed these parameters and found that myriocin did not significantly reduce plasma cholesterol or TG concentrations (Table 1) or appear to modify lipoprotein profile (Fig. 4); in general agreement with Hojjati et al. [22]. When we analysed plasma SM concentrations, we

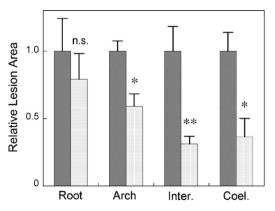


Fig. 3 – Morphometric analysis of atherosclerotic lesions. ApoE $^{-/-}$ mice were fed a high fat diet for 63 days and concurrently treated i.p. with myriocin at 0.3 mg/kg/48 h i.p. or phosphate buffered saline as vehicle control. Lesion areas were measured in myriocin (dark bars) and PBS (light bars) treated mice (n=7 in each group) at the aortic root (Root), arch (Arch) third intercostal branch (Inter.) and coeliac branch (Coel.). All PBS control conditions were assigned a value of 1.0 and relative changes in lesion area induced by myriocin are shown. Data are means \pm S.E. $^{\circ}$ P < 0.05, $^{\circ}$ P < 0.01 assessed by Mann–Whitney U test.

found that myriocin treatment resulted in a significant 42% decrease overall which was not as profound as the 59% reduction in plasma SM previously reported [22]. Nonetheless, this result does confirm that highly significant reductions in plasma SM concentration are achievable in the absence of overt changes to other potentially atherogenic lipids (cholesterol and TG) or lipoprotein profile.

The primary aim of the present study was to examine potential changes in plasma GSL concentration that may be induced with myriocin treatment. In order to examine this we analysed GSL glycans by HPLC; a method which we have previously shown is a quantitative and sensitive enough for analysis of individual mouse plasma samples [10,29]. In agreement with our previous work, LacCer, GalNAc β 1-1Gal β 1-4Glc-Cer (GA2) and N-glycolyl GalNAc β 1-4[NeuNGc α 2-3]Gal β 1-4Glc-Cer (gGM2) represented the three major plasma GSL identified (Fig. 5A). In the mice treated with

Table 1 – Impact of SPT inhibition on plasma lipid concentrations			
TC (mM)	TG (mM)	SM (mM)	GSL (μM)
PBS 25.5 (2.0)	1.24 (0.30)	1.94 (0.14)	68.9 (3.8)
MYR 27.9 (3.4) n.s.	1.81 (0.35) n.s.	1.13 (0.08) 0.0004	51.7 (4.4) 0.0143

ApoE $^{-/-}$ mice fed a high fat diet were injected i.p. every 48 h with 100 μ l phosphate buffered saline (PBS) or PBS containing myriocin 0.3 mg/kg (MYR) for 63 days. Plasma lipids were then determined as described in Section 2 after an over-night fast. Values are means (n=7) with S.E. in parentheses and level of significance (Student's t-test, P-value) in italics. n.s., not significant.

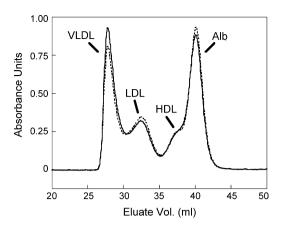


Fig. 4 – Analysis of plasma lipoprotein profile by FPLC. ApoE^{-/-} mice were fed a high fat diet for 63 days and concurrently treated i.p. with myriocin (solid line) at 0.3 mg/kg/48 h i.p. or phosphate buffered saline (broken line) as vehicle control. Plasma samples were analysed by FPLC and the elution positions of the major lipoprotein fractions and albumin assessed by monitoring absorbance at 280 nm. VLDL, very low density lipoprotein; LDL, low density lipoprotein; HDL, high density lipoprotein; Alb, albumin.

myriocin, a significant 25% reduction in total GSL concentration (LacCer + GA2 + gGM2) was observed (Table 1). Furthermore, plasma SM and GSL concentrations were significantly correlated (Fig. 5B). This data therefore suggests that the inhibition of atherosclerosis induced by myriocin may be due to reductions in both plasma SM and GSL concentrations.

4. Discussion

Previous studies showed that myriocin administered either orally (0.3 mg/kg/day) or i.p. (0.3 mg/kg/48 h) significantly reduced atherosclerosis in apo $E^{-/-}$ mice [21,22]. These studies both suggested that reduced plasma SM levels were responsible for the inhibition of atherosclerosis; however, it was not clear if additional atherogenic lipids may be altered with myriocin treatment. Based on the knowledge that myriocin administered via the i.p. route inhibits hepatic SPT activity by 50% [22], that SPT catalyses the first step in the GSL biosynthetic pathway (Fig. 6), and that the vast majority of plasma GSL are derived from the liver [32], we predicted that plasma GSL levels may also be reduced with myriocin treatment. We used an i.p. administration route to avoid the potentially confounding antiatherogenic reductions in plasma cholesterol and TG concentrations and alterations in plasma lipoprotein profile that result from oral administration [31]. Our data indicate that reductions in both plasma SM and GSL concentrations may play a role in the anti-atherogenic mechanisms associated with myriocin. The development of more selective GSL synthesis inhibitors and methods to quantify both inhibitors and GSLs in distinct lesion sites would obviously help to resolve the specific contributions of GSL to atherosclerosis.

Another novel observation in the present study concerns the site-specific inhibition of lesions in mice treated with myriocin.

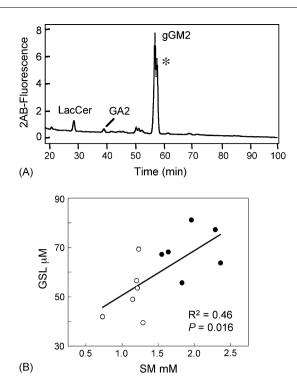


Fig. 5 – Analysis of plasma GSL concentration by HPLC and correlation of GSL and SM concentrations. (A) Plasma GSL from a control PBS treated mouse were subjected to ceramide glycanse digestion and the resulting glycans labelled with 2-aminobenzamide (2-AB) and analysed by normal phase HPLC. The elution positions of lactosyl ceramide (LacCer), GalNAc β 1-1Gal β 1-4Glc-Cer (GA2) and N-glycolyl GalNAc β 1-4[NeuNGc α 2-3]Gal β 1-4Glc-Cer (gGM2) are shown. The asterisk denotes previously identified gGM2 lactone [10]. (B) Correlation of plasma GSL and SM concentrations. Myriocin treated (\bigcirc), PBS treated (\bigcirc).

We detected more potent inhibition of atherosclerotic lesions with increasing distance from the heart (Fig. 3). While the mechanisms responsible for this site-specific action are unclear, this may be a general phenomenon associated with pharmacological treatment of apo $E^{-/-}$ mice since the antiatherogenic actions of vitamin E plus coenzyme Q_{10} [33] as well as probucol [26] are also more potent at the distal lesion sites.

While speculative, it seems reasonable to propose that targeting of SM and/or GSL synthesis could offer a novel pharmacological approach to treat human atherosclerosis that may be complementary to lowering total plasma cholesterol concentrations [24]. This could represent a new therapeutic approach for patients who do not respond well to statin treatment and opens up opportunities for a dual treatment approach (i.e. cholesterol and SM/GSL synthesis inhibition) to treating CVD in the population at large.

In conclusion, we have shown that myriocin inhibits the development of atherosclerosis in apoE^{-/-} mice fed a high fat diet and that the anti-atherogenic effect is more pronounced at distal sites of the aorta. The inhibition of atherosclerosis was not associated with reductions in plasma cholesterol or TG concentration or with altered lipoprotein profile but was

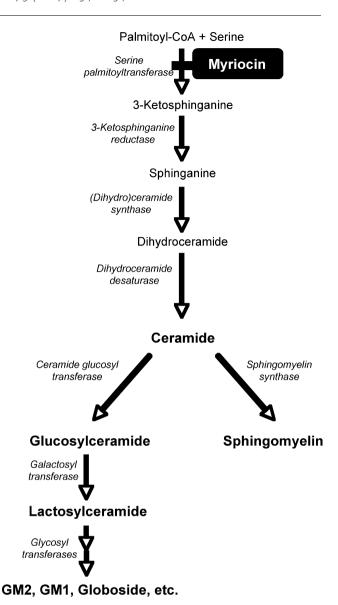


Fig. 6 – Simplified overview of the sphingolipid biosynthetic pathway. Serine palmitoyltransferase catalyses the initial step in both glycosphingolipid (e.g. lactosylceramide, GM1, GM2, globoside) and sphingomyelin biosynthesis and our data indicate that the in vivo inhibition of this enzyme by myriocin results in a reduction of both these sphingolipid classes.

associated with significant reductions in plasma SM and GSL concentrations. These data suggest that therapeutic reduction of plasma GSL concentration may contribute to the previously reported anti-atherosclerotic actions associated with SPT inhibition.

REFERENCES

- [1] Lusis AJ. Atherosclerosis. Nature 2000;407:233-41.
- [2] Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. N Engl J Med 2005;352:1685–95.

- [3] Libby P, Ridker PM, Maseri A. Inflammation and atherosclerosis. Circulation 2002;105:1135–43.
- [4] Stocker R, Keaney Jr JF. Role of oxidative modifications in atherosclerosis. Physiol Rev 2004;84:1381–478.
- [5] Smith EB. Intimal and medial lipids in human aortas. Lancet 1960;1:799–803.
- [6] Breckenridge WC, Halloran JL, Kovacs K, Silver MD. Increase of gangliosides in atherosclerotic human aortas. Lipids 1975;10:256–9.
- [7] Prokazova NV, Bergelson LD. Gangliosides and atherosclerosis. Lipids 1994;29:1–5.
- [8] Mukhin DN, Chao FF, Kruth HS. Glycosphingolipid accumulation in the aortic wall is another feature of human atherosclerosis. Arterioscler Thromb Vasc Biol 1995;15:1607–15.
- [9] Jiang XC, Paultre F, Pearson TA, Reed RG, Francis CK, Lin M, et al. Plasma sphingomyelin level as a risk factor for coronary artery disease. Arterioscler Thromb Vasc Biol 2000:20:2614-8.
- [10] Garner B, Priestman DA, Stocker R, Harvey DJ, Butters TD, Platt FM. Increased glycosphingolipid levels in serum and aortae of apolipoprotein E gene knockout mice. J Lipid Res 2002;43:205–14.
- [11] Garner B, Mellor HR, Butters TD, Dwek RA, Platt FM. Modulation of THP-1 macrophage and cholesterol-loaded foam cell apolipoprotein-E levels by glycosphingolipids. Biochem Biophys Res Commun 2002;290:1361–7.
- [12] Glaros EN, Kim WS, Quinn CM, Wong J, Gelissen I, Jessup W, et al. Glycosphingolipid accumulation inhibits cholesterol efflux via the ABCA1/apoA-I pathway. 1-Phenyl-2-decanoylamino-3-morpholino-1-propanol is a novel cholesterol efflux accelerator. J Biol Chem 2005;280:24515–23.
- [13] Gong N, Wei H, Chowdhury SH, Chatterjee S. Lactosylceramide recruits PKCalpha/epsilon and phospholipase A2 to stimulate PECAM-1 expression in human monocytes and adhesion to endothelial cells. Proc Natl Acad Sci USA 2004;101:6490–5.
- [14] Bhunia AK, Han H, Snowden A, Chatterjee S. Redoxregulated signaling by lactosylceramide in the proliferation of human aortic smooth muscle cells. J Biol Chem 1997;272:15642–9.
- [15] Wen FQ, Jabbar AA, Patel DA, Kazarian T, Valentino LA. Atherosclerotic aortic gangliosides enhance integrinmediated platelet adhesion to collagen. Arterioscler Thromb Vasc Biol 1999;19:519–24.
- [16] Moon SK, Kang SK, Kim CH. Reactive oxygen species mediates disialoganglioside GD3-induced inhibition of ERK1/2 and matrix metalloproteinase-9 expression in vascular smooth muscle cells. FASEB J 2006;20:1387–95.
- [17] Levade T, Auge N, Veldman RJ, Cuvillier O, Negre-Salvayre A, Salvayre R. Sphingolipid mediators in cardiovascular cell biology and pathology. Circ Res 2001;89:957–68.
- [18] Oram JF, Lawn RM. ABCA1. The gatekeeper for eliminating excess tissue cholesterol. J Lipid Res 2001;42:1173–9.
- [19] Rader DJ. Regulation of reverse cholesterol transport and clinical implications. Am J Cardiol 2003;92:42J–9J.

- [20] Bodary PF, Shen Y, Vargas FB, Bi X, Ostenso KA, Gu S, et al. Alpha-galactosidase A deficiency accelerates atherosclerosis in mice with apolipoprotein E deficiency. Circulation 2005:111:629–32.
- [21] Park TS, Panek RL, Mueller SB, Hanselman JC, Rosebury WS, Robertson AW, et al. Inhibition of sphingomyelin synthesis reduces atherogenesis in apolipoprotein E-knockout mice. Circulation 2004;110:3465–71.
- [22] Hojjati MR, Li Z, Zhou H, Tang S, Huan C, Ooi E, et al. Effect of myriocin on plasma sphingolipid metabolism and atherosclerosis in apoE-deficient mice. J Biol Chem 2005:280:10284–9.
- [23] Miyake Y, Kozutsumi Y, Nakamura S, Fujita T, Kawasaki T. Serine palmitoyltransferase is the primary target of a sphingosine-like immunosuppressant, ISP-1/myriocin. Biochem Biophys Res Commun 1995;211:396–403.
- [24] Kim WS, Chalfant CE, Garner B. Fine tuning therapeutic targeting of the sphingolipid biosynthetic pathway to treat atherosclerosis. Current Vasc Pharmacol 2006;4:151–4.
- [25] Mach F, Schonbeck U, Sukhova GK, Atkinson E, Libby P. Reduction of atherosclerosis in mice by inhibition of CD40 signalling. Nature 1998;394:200–3.
- [26] Witting PK, Pettersson K, Letters J, Stocker R. Site-specific antiatherogenic effect of probucol in apolipoprotein Edeficient mice. Arterioscler Thromb Vasc Biol 2000;20:E26– 33
- [27] Choy K, Beck K, Png FY, Wu BJ, Leichtweis SB, Thomas SR, et al. Processes involved in the site-specific effect of probucol on atherosclerosis in apolipoprotein E gene knockout mice. Arterioscler Thromb Vasc Biol 2005;25:1684–90.
- [28] Hojjati MR, Jiang XC. Rapid, specific, and sensitive measurements of plasma sphingomyelin and phosphatidylcholine. J Lipid Res 2006;47:673–6.
- [29] Wing DR, Garner B, Hunnam V, Reinkensmeier G, Andersson U, Harvey DJ, et al. High-performance liquid chromatography analysis of ganglioside carbohydrates at the pmol level after ceramide glycanse digestion and fluorescent labelling with 2-aminobenzamide. Anal Biochem 2001;298:207–17.
- [30] Suarna C, Wu BJ, Choy K, Mori T, Croft K, Cynshi O, et al. Protective effect of vitamin E supplements on experimental atherosclerosis is modest and depends on preexisting vitamin E deficiency. Free Radic Biol Med 2006;41:722–30.
- [31] Park TS, Panek RL, Rekhter MD, Mueller SB, Rosebury WS, Robertson A, et al. Modulation of lipoprotein metabolism by inhibition of sphingomyelin synthesis in ApoE knockout mice. Atherosclerosis 2006;189:264–72.
- [32] Senn HJ, Sellin S, Fitzke E, Stehle T, Haussinger D, Wieland H, et al. Biosynthesis and excretion of gangliosides by the isolated perfused rat liver. Eur J Biochem 1992;205:809–14.
- [33] Thomas SR, Leichtweis SB, Pettersson K, Croft KD, Mori TA, Brown AJ, et al. Dietary cosupplementation with vitamin E and coenzyme Q(10) inhibits atherosclerosis in apolipoprotein E gene knockout mice. Arterioscler Thromb Vasc Biol 2001;21:585–93.